the dinitrobenzenes, however, electron transfer can only be observed photochemically. Based on our observation of photochemical oxidation of Mo(NCS)₆³⁻ by 1,3-dinitrobenzene, the redox-active $Mo(NCS)_6^{3-}$ excited state may have an energy as high as 1.3 eV. Since the $Mo(NCS)_6^{3-}$ absorption spectrum indicates that the lowest energy excited state lies no higher than 1.0 eV, our photochemical experiments suggest that upper excited states of $Mo(NCS)_6^{3-}$ may be participating in these electrontransfer reactions.

The properties of photogenerated Mo(NCS)₆²⁻ can be explored further by using CCl_4 as an irreversible electron acceptor. One-electron reduction of CCl₄ is followed by rapid fragmentation;¹² thus, back electron-transfer reactions such as (2) should be inhibited. Flash irradiation of $Mo(NCS)_6^{3-}$ in CH₃CN-CCl₄ (90:10 w/w) results in immediate permanent formation of redbrown $Mo(NCS)_6^{2-}$, which is stable under these conditions.

Photooxidation of $Mo(NCS)_6^{3-}$ (ca. 8 × 10⁻³ M; 0.2 M KNCS) is also observed in CCl₄-H₂O-CH₃CN (7:31:62 w/w; chosen to keep H_2O and CCl_4 stable in the same phase), but the Mo- $(NCS)_{6}^{2-}$ that is produced slowly disappears. After several flashes, each of which leads to formation and slow disappearance of $Mo(NCS)_6^{2-}$, a new, permanent absorbance at 456 nm is observed. We attribute this new feature to the molybdenum(V) product formed by disproportionation of $Mo(NCS)_6^{2-}$. Of the three possible Mo(V) products suggested by Sykes and co-workers,⁸ both $MoO(NCS)_5^{2-13}$ and $Mo_2O_4(NCS)_6^{4-10}$ exhibit intense absorption near 456 nm. However, the mononuclear complex is stable only in strongly acidic solutions.^{13b,14} Thus, $Mo_2O_4(NCS)_6^{4-}$ is the most likely product under our conditions, and the overall sequence of reactions is as follows:

$$Mo(NCS)_6^{3-} + CCl_4 \xrightarrow{n\nu} Mo(NCS)_6^{2-} + [CCl_4^{-}]$$
 (3)

$$[CCl_4^-] \xrightarrow{fast} decomposition products$$
 (4)

 $4M_0(NCS)_6^{2^-} + 4H_2O \xrightarrow{slow} 2M_0(NCS)_6^{3^-} + (SCN)_3M_0O(\mu-O)_2M_0O(NCS)_3^{4^-} + 8H^+ + 6NCS^- (5)$

We observe first-order kinetics $(k_{obsd} \text{ ca. 7 s}^{-1})$ for disappearance of $Mo(NCS)_6^{2-}$; this is consistent with a mechanism for reaction 5 that involves initial rate-limiting aquation of $Mo(NCS)_6^{2-}$.

We have therefore shown for the first time that molybdenum-(III) complexes can undergo facile photoredox reactions and that photoinitiated two-electron oxidations can be accomplished by disproportionation of the initial electron-transfer products.¹⁵ Experiments now in progress include nanosecond flash photolysis¹⁶ and selective irradiation of $Mo(NCS)_6^{3-}$ in its weak low-energy absorption bands, in order to examine the redox-active excited states more directly. These results are of particular interest because of the possibility that electron transfer may be occurring from upper excited states. Also under study are the extension of these reactions to other second- and third-row d³ complexes and the reactivity of the photogenerated oxo complexes toward organic and inorganic substrate molecules.

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Stereochemistry of Methylation in Thienamycin Biosynthesis: Example of a Methyl Transfer from Methionine with Retention of Configuration

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The carbapenem antibiotic thienamycin¹ (I) is derived biosynthetically from glutamic acid, which provides the pyrroline ring,² acetate and cysteine, providing C-6 and C-7 of the β -lactam ring and the cysteamine side chain, respectively,³ and the methyl group of methionine, which gives rise to both carbon atoms of the hy-droxyethyl side chain.³ In a double-labeling experiment, Wil-liamson et al.³ demonstrated that [methyl-¹⁴C,³H]methionine is incorporated with 58% tritium retention, relative to ¹⁴C, corresponding to 87% of the maximum value for retention of four of the six hydrogens of the two methyl groups.

In order to obtain more information on this intriguing double-methylation sequence, we examined the steric course of the transfer of the methionine methyl group. We first confirmed the incorporation of an intact methyl group into C-9 by using L-[methyl- ${}^{13}C, {}^{2}H_{3}$] methionine. This material, synthesized⁴ from ${}^{13}C^{2}H_{3}$ I (Merck, 99% ${}^{13}C$, 98% ${}^{2}H$), was fed at a concentration of 0.74 mM to 1.2 L (3 \times 40 mL) of a resting cell suspension of Streptomyces cattleya.³ The fermentation was terminated 19 h later and I (3.2 mg) was isolated as described earlier.^{3,5} The ¹³C NMR spectrum of the sample was recorded at 4 °C at 75.5 MHz (Bruker WM 300) with proton broad-band decoupling and with and without deuterium broad-band decoupling. The results (Figure 1) clearly show that ¹³C is incorporated into C-9 with retention of all three deuterium atoms and into C-8 with retention of one atom of deuterium. There seems to be little or no washout of deuterium from either position.

With this information at hand, we fed (methyl-R)- and (methyl-S)-[methyl-²H₁,³H]-L-methionine⁶ (R, 2.79 mCi/mmol, max 80% ee methyl-R; S, 1.10 mCi/mmol, max 75% ee methyl-S⁷) to 80-mL cultures of S. cattleya (15 μ Ci per experiment). When tritium incorporation into I had reached a maximum, as determined by HPLC and liquid scintillation counting, 2 mg of unlabeled I was added and the labeled I was isolated and purified^{3,5} (1-4% tritium incorporation). Kuhn-Roth oxidation of the products gave acetic acid samples which were analyzed for their chirality by the method of Cornforth⁸ and Arigoni⁹ using our previously described procedure.¹⁰ The acetic acid obtained by

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¹²⁵⁵

⁽⁷⁾ The (R)- and (S)-methionine samples were synthesized from chiral acetic acid of F = 20 and 78, respectively. Since the first step in the reaction sequence, the Schmidt reaction,⁶ can involve some racemization, we analyzed the chiral purity of the resulting methionine by conversion to idolmycin fol-lowed by Kuhn-Roth oxidation to acetic acid.⁶ The values for the chiral purity of the methionine sample correspond to those measured for the methioninederived acetate.

Scheme I



¹³C NMR spectrum of thienamycin biosynthesized from Figure 1. [methyl-13C, 2H3] methionine. Signals for C-8 (left) and C-9 (right) with broad-band ¹H decoupling (top tracings) and with broad-band ¹H and ²H decoupling (bottom tracings).

degradation of I from (methyl-R)-methionine had an F value¹⁰ of 69 indicating 66% ee R configuration of the thienamycin methyl group. Conversely, the acetate from the (methyl-S)-methionine experiment had F = 35 corresponding to 52% ee S configuration of the thienamycin methyl group. A complete repetition of the experiments confirmed these results.

The above results demonstrate that the biosynthesis of thienamycin involves transfer of a methyl group from methionine or, most likely, its activated form, S-adenosylmethionine¹¹ (AdoMet), to an acceptor carbon with net retention of configuration (Scheme This is in marked contrast to the findings with all other D. AdoMet-dependent methyltransferases studied to date which, without exception, have been found to catalyze methyl transfer with inversion of configuration. The 11 examples include enzymes catalyzing methyl transfer to oxygen, ¹²⁻¹⁴ nitrogen, ^{6,13,15,16} and sulfur¹³ as well as to carbon. ^{6,13,17} This stereochemical uniformity has been interpreted to indicate a single transfer of the methyl group directly from the sulfur of AdoMet to the acceptor nucleophile in an $S_N 2$ transition state.¹² Methyl transfer to C-8 of I could be followed by hydride migration from C-8 to C-6 and proton loss from the methyl group to give a C-9 methylene group, in analogy to the side-chain methylation of sterols.¹³ Such a

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methylene group could be reduced to give either an R or an Smethyl group. However, the full retention of three atoms of deuterium from [13C,2H3]methionine at C-9 argues against this possibility. It seems more likely, then, that C-9 of I is derived by a mechanistically different methylation reaction involving two sequential methyl transfers, each proceeding with inversion. Since the thienamycin fermentation has been noted to have a cobalt requirement,³ one may tentatively speculate that a corrin could serve as an intermediate carrier. This would have an analogy in the formation of methionine from 5-methyltetrahydrofolate by B12-dependent methionine synthase, which we have recently shown to proceed with net retention of methyl group configuration.¹⁸

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Enhancement of Radioactivity of ¹⁴C-¹²C Mixtures via **Partial Reduction**

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Observable nuclear transmutation reactions have occupied an extremely important position in nuclear medicine, nuclear energy, and geological and anthropological dating. Further, the concentration of radioactive materials (enhancement of radioactivity) has grown to be very important in the treatment and storage of radioactive waste and in the development of techniques for reactivating spent nuclear fuel. These facts coupled with our observation¹ that the solution electron affinities of perdeuterated polyaromatics are less than that of the protiated materials led us to investigate the possibility of increasing the radioactivity of benzophenone-carbonyl- ^{14}C (BZO-14C)-cold benzophenone (BZO-12C) mixtures via the partial reduction of these mixtures to the ketyls.

When 0.6-5.0-mmol samples of a radioactive $(0.02-0.16 \ \mu \text{Ci})^2$ mixture of BZO-14C and BZO-12C are reduced with deficient amounts of sodium metal in liquid ammonia (20-40 mL), the concentrations of the cold and hot anion radicals and neutral molecules are controlled via reaction 1.

 $BZO-12C^{-} + BZO-14C \Rightarrow BZO-12C + BZO-14C^{-} (1)$

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